Low-Density Lipoprotein Cholesterol (LDL-C) Content Assay Kit

Note: It is necessary to predict 2-3 large difference samples before the formal determination.

Operation Equipment: Spectrophotometer/microplate reader

Catalog Number: NA0146

Size: 100T/96S

Components:

Extract: Isopropyl alcohol 110 mL ×1. Required but not provided. Store at 2-8°C.

Reagent IA: Liquid 25 mL×1. Store at 2-8°C. **Reagent IB:** Liquid 200 μ L×1. Store at 2-8°C.

Reagent IC: Liquid 30 μL×1. Store at 2-8°C.

Reagent I: According to the ratio of Reagent IA: Reagent IB: Reagent IC=2.25 mL: 20 μ L: 3 μ L (about 12T) to prepare when the solution will be used.

Reagent IIA: Powder ×2. Store at 2-8°C.

Reagent IIB: Liquid 10 mL×1. Store at 2-8°C. Add 4mL of Reagent IIB to one Reagent IIA and shake to dissolve. It could be stored at 2-8°C for one week.

Standard Solution: Powder $\times 1$, 10 mg cholesterol. Store at 2-8°C. Add 517 μ L of Extract before use and shake to dissolve. The cholesterol standard solution of 50 μ mol/mL could be stored at 2-8°C for four weeks.

Product Description

Low-density lipoproteins (LDL) are the major carriers of cholesterol in humans, responsible for supplying cholesterol to tissues with the highest sterol demands. Low-density lipoprotein cholesterol (LDL-C) concentrations positively correlate with the incidence of coronary heart disease and a reduction of LDL-C decreases the risk of coronary. Therefore, accurate and precise measurements of patients' LDL-C concentrations are necessary to appropriately identify individuals with atherosclerosis, coronary heart disease and hypertension.

Cholesterol of chylomicrons (CM), very-low-density lipoproteins (VLDL), high-density lipoproteins (HDL) is specifically dissociated by one surfactant, but LDL-C is not dissociated by the surfactant. Cholesterol ester and cholesterol oxidase can catalyze the hydrolysis of dissociated cholesterol to produce H_2O_2 , which cannot form colored compounds without chromogenic agents. Cholesterol is specifically dissociated by another surfactant from undissociated LDL. Esterase can catalyze the hydrolysis of cholesterol ester to produce free cholesterol (FC) and free fatty acid (FFA), thus transforming cholesterol ester into FC; Furthermore, cholesterol oxidase can catalyze FC to form $\Delta 4$ -cholesterone and H_2O_2 ; Finally, peroxidase can catalyze the oxidation of 4-aminoantipyrine and phenyl amines by H_2O_2 to form purple quinones. It has a characteristic absorption peak at 546 nm, and its color depth is directly proportional to cholesterol content.

Reagents and Equipment Required but Not Provided.

Spectrophotometer/microplate reader, balance, low temperature table centrifuge, constant temperature incubator/water bath, micro glass cuvette/96 well flat-bottom plate, pipette, mortar/homogenizer/cell ultrasonic crusher, ice, distilled water, **isopropyl alcohol**.

Procedure

I. Sample preparation:

- **1. Tissue:** according to the tissue weight (g): the Extract volume (mL) is 1:5-10. (It is recommended that add 1 mL of Extract to 0.1 g tissue). Homogenate in ice bath, then centrifuge at 10000 g for 10 minutes at 4°C. Take the supernatant for test.
- **2. Bacteria/cells:** according to the number of bacteria/cells (10⁴): the volume of Extract (mL) is 500~1000:1. It is suggest that add 1 mL of Extract to 500 million of cells. Breaking bacteria/cells by ultrasonic wave in ice bath (power 300W, ultrasonic 2s, interval 3s, total time 3 min). Centrifuge at 10000 g 4°C for 10 minutes. Take the supernatant on ice for test.
- **3. Serum (plasma) or other liquid samples:** detect directly. Centrifuge before detecting if there are precipitation in the liquid.

II. Determination Procedure

- 1. Preheat the spectrophotometer/microplate reader for more than 30 minutes, adjust the wavelength to 546 nm and set spectrophotometer counter to zero with distilled water.
- 2. Standard working solution: Dilute 50 μ mol/mL standard solution with distilled water to 2.5, 1.25, 0.625, 0.3125, 0.15625, 0.078125, 0.0390625 μ mol/mL for standby.

3. Operation table:

Reagent (µL)	Test tube (A _T)	Standard tube (A _S)	Blank tube (A _B)
Sample	5	-	-
Standard	-	5	-
Extract	-	-	5
Reagent I	180	180	180
Mix well. React at 37°C for 5 minutes. Measure the absorption at 546 nm and record as A1 _T , A1 _S , A1 _B .			
Reagent II	60	60	60

Mix thoroughly. React at 37°C for 5 minutes. Measure the absorption at 546 nm and record as A2_T, A2_S, A2_B. Calculate Δ A_T = (A2_T - A1_T)- (A2_B - A1_B), Δ A_S = (A2_S - A1_S)- (A2_B - A1_B). Blank tube and standard curve only need to test once or twice.

III. Calculation of LDL-C Content:

1. Standard curve

Taking the concentration of each standard solution as the x-axis and its corresponding ΔA_S as the y-axis, draw a standard curve to get the standard equation y = kx + b, and bring ΔA_T into the equation to get x (μ mol/mL).

2. Calculation

- 1) Serum (plasma) or other liquid samples: LDL-C content (μ mol/dL) =x×100
- 2) Protein concentration: LDL-C content (μ mol/mg prot) = $x \times V_S \div (Cpr \times V_S) = x \div Cpr$
- 3) Sample weight: LDL-C content (μ mol/g weight) = $x \times Vs \div (W \div V_E \times Vs) = x \div W$
- 4) Bacteria/cells number: LDL-C content (nmol/10⁴ cell) = $x \times Vs \div (500 \div V_E \times Vs) \times 10^3 = 2x$

100: Unit conversion factor, 1 dL=100 mL;

V_S: Added sample volume, 0.005 mL;

V_E: Extract volume, 1 mL;

W: Sample weight, g;

500: The number of bacteria/cells, 500 million;

Cpr: The concentration of protein, mg/mL;

10³: Unit conversion factor, 1 μmol=10³ nmol.

Note:

- 1. If samples ΔA_T is too high, it is suggested that the samples should be diluted with multiple times of Extract solution. Sample supernatant volume could be increased if samples ΔA_T is too low. And modify the calculation formula.
- 2. The protein concentration can be detected in another tissue.

Experimental example:

- 1. Take 5 μ L of human serum, operate according to the determination steps, calculate $\Delta A_T = (A2_T A1_T) (A2_B A1_B) = (0.515 0.061) (0.056 0.055) = 0.453$. Bring the result into the standard curve y=0.1163x-0.0039 and calculate x=3.929. The result is calculated according to liquid volume: LDL-C content (μ mol/dL) =x×100=3.929×100=392.863 μ mol/dL.
- 2. Take 0.11g mice liver, add 1 mL of Extract, grind the homogenate with ice bath. Then operate according to the determination steps, calculate $\Delta A_T = (A2_T A1_T) (A2_B A1_B) = (0.239 0.056) (0.056 0.055) = 0.182$. Bring the result into the standard curve y=0.1163x-0.0039 and calculate x=1.598. The result is calculated according to sample weight:
 - LDL-C content (μ mol/g weight) = $x \div W = 1.598 \div 0.11 = 14.531 \ \mu$ mol/g weight.

References:

- [1] Hiroyuki S, Tetsumi I, Yoshinori U, et al. Homogeneous assay for measuring low-density lipoprotein cholesterol in serum with triblock copolymer and α-cyclodextrin sulfate[J]. Clinical Chemistry, 1998, 44(3):522-531.
- [2] Sakaue T, Hirano T, Yoshino G, et al. Reactions of direct LDL-cholesterol assays with pure LDL fraction and IDL: comparison of three homogeneous methods[J]. Clinica Chimica Acta, 2000, 295(1-2):97-106.

Related products:

NA0822/NA0580	Free fatty Acids(FFA) Content Assay Kit
NA0808/NA0566	Acetaldehyde Dehydrogenase(ALDH) Activity Assay Kit
NA0733/NA0491	Free Cholestenone(FC) Content Assay Kit
NA0727/NA0485	Total Cholestenone(TC) Content Assay Kit
NA0153/NA0152	High-Density Lipoprotein Cholesterol (HDL-C) Content Assay Kit