

## D-Xylose Content Assay Kit

**Note:** Take two or three different samples for prediction before test.

**Operation Equipment:** Spectrophotometer

**Catalog Number:** NA0294

**Size:**50T/48S

### Components:

Extract solution: 55mL×1, storage at 4°C.

Reagent 1A: Powder×4, storage at 4°C.

Reagent 1B: 80mL×1, storage at 4°C.

Standard: Powder×1, storage at 4°C. 10mg of D-xylose. Add 1 mL of distilled water to prepare a 10 mg / mL xylose standard solution.

Preparation of Reagent 1: Add 20mL of Reagent1B to each Reagent 1A before use and dissolve. If the reagent discolors, it has deteriorated.

### Product Description:

Xylose is a type of pentose. Natural D-xylose exists in plants in the form of polysaccharides. Xylose can activate and promote the growth of bifidobacteria in the human intestinal tract. Bifidobacteria are beneficial bacteria. The more bacteria, the more beneficial to human health. Eating xylose can improve the human microbial environment and improve the immune system of the body. After being absorbed in the upper small intestine, sugar does not participate in metabolism in the body and is excreted by the kidneys. Therefore, the absorption of D-xylose has been an important functional indicator of small intestinal malabsorption.

D-xylose is hydrolyzed to produce furfural under strong acid conditions. Furfural can react with resorcinol to form pink compounds. It has a special absorption peak at 554 nm. Based on this, the content of xylose in the sample can be calculated from the absorbance.

### Reagents and Equipment Required but Not Provided:

Spectrophotometer, water-bath/constant temperature incubator, adjustable transferpettor, mortar/homogenizer, centrifuge, 30-50 mesh sieve, 1 mL glass cuvette, [EP tube](#) and distilled water.

### Sample preparation:

1. Plant samples: The plant samples are dried in a blast oven at 65°C, ground into a powder, and pass through a 30-50 mesh sieve. According to weight (g): volume of extract solution (mL)= 1: 50 ~ 100 ratio (we recommend to weigh 20mg of dry sample and add 1.0 mL of extract solution), vortex to mix, hydrolyze in 100°C water bath for 2 h, then centrifuge [at 10000rpm and room temperature](#) for 15 min, discard the pellet, and take the supernatant for testing;
2. Other tissue: According to weight (g): volume of extract solution (mL)= 1: 5~10 ratio (we recommend

to weigh 0.1g sample and add 1.0 mL extract solution), homogenize in ice bath, and hydrolyze in a 100°C water bath 2 h, then centrifuge at 4°C, 10,000 rpm for 15 min, discard the precipitate, and take the supernatant on ice for testing;

3. Serum (plasma) samples: directly test.

**Procedure:**

1. Preheat spectrophotometer for 30min, adjust the wavelength to 554 nm and set the counter to zero with distilled water.
2. Standard solution: The standard is diluted with distilled water to 0.4, 0.25, 0.125, 0.0625, 0.03125, 0.015625, 0.0078mg/mL.
3. Operation table: (operation in 1.5mL EP tube):

Reagent name (μL)	Test tube (A <sub>T</sub> )	Standard tube (A <sub>S</sub> )	Blank tube (A <sub>B</sub> )
Sample	200		
Standard solution		200	
Distilled water			200
Reagent 1	1000	1000	1000

After mixing, boil in a water bath for 8 minutes, then cool to room temperature in an ice bath and measure the absorbance at 554 nm. Record the results as A<sub>T</sub>, A<sub>S</sub>, and A<sub>B</sub>. calculate  $\Delta A_T = A_T - A_B$ ,  $\Delta A_S = A_S - A_B$ ,

**Calculation:**

1. Taking the concentration of each standard solution as the x-axis and its corresponding  $\Delta A_S$  as the y-axis, draw a standard curve to get the standard equation  $y = kx + b$ , and bring  $\Delta A_T$  into the equation to get x (mg / mL)

2. **Plant sample:**

$$D\text{-xylose (mg/g sample)} = V_e \times x \div W_1 = 0.5x \div W_1$$

3. **Other sample:**

$$D\text{-xylose (mg/g sample)} = V_e \times x \div W_2 = 0.5x \div W_2$$

4. **Liquid volume:**

$$D\text{-xylose (mg/mL)} = V_s \times x \div V_s = x$$

V<sub>s</sub>: sample volume, 0.2mL;

V<sub>e</sub>: volume used in the extraction solution, 1mL;

W: Fresh weight of sample, g;

**Note:**

1. The same batch of test samples need to be equipped with 1-2 blank tubes, standard tubes need only be tested 1-2 times.
2. When the A is higher than 0.9, it is recommended to test the sample after dilution and multiply it by the

dilution factor in the calculation formula.

3. Reagent 1B has a strong irritating odor. Excessive inhalation is harmful to the human body, so it is recommended to operate in a fume hood.

#### **Experimental Examples:**

1. Take 0.05g of Euonymus stem , and add 1 mL of extract solution for sample processing, take the supernatant and dilute 2 times according to the determination procedure, measure and calculate  $\Delta A = A_t - A_b = 0.639 - 0.002 = 0.637$ , Bring in the standard curve  $y = 2.3529x + 0.0015$ , calculate  $x = 0.27$ , and calculate according to the formula:

D-Xylose Content (mg/g) =  $x \times V_{\text{extraction}} \div W_1 \times 2$  (dilution multiple) =  $0.27 \times 0.5 \div 0.05 \times 2 = 5.4$  mg/g.

#### **Related Products:**

NA0681/NA0439  $\beta$ -xylosidase Activity Assay Kit

NA0675/NA0433 Glucose Dehydrogenase(GCDH) Activity Assay Kit

NA0312/NA0311 N-Acetyl- $\beta$ -D-Glucosidase(NAG) Activity Assay Kit

#### **Technical Specifications :**

Minimum Detection Limit: 0.0007 mg/mL

Linear Range: 0.0039-0.4 mg/mL