# Glycogen Phosphorylase a (GPa) Activity Assay Kit

Note: Take two or three different samples for prediction before test.

**Operation Equipment:** Ultraviolet spectrophotometer/Microplate reader

Catalog Number: NA0390

Size: 100T/96S

#### **Components:**

Reagent	Size	Storage
Extract solution	110 mL×1	4°C
Reagent I	20 mL×1	4°C
Reagent II	Powder×1	4°C
Reagent III	Powder×1	4°C
Reagent IV	Powder×1	-20°C
Reagent V	Powder×2	-20°C
Reagent VI	Powder×2	-20°C

Solution preparation:

1. Reagent II: Dissolved with 0.5 mL of distilled water before use. Mix thoroughly.

2. **Reagent III:** Dissolved with 0.5 mL of distilled water before use. Mix thoroughly. It can be divided into small tubules and stored for 4 weeks at -20°C. Avoid repeating freeze/thaw cycles.

3. **Reagent IV:** Dissolved with 1.25 mL of distilled water before use. Mix thoroughly. It can be divided into small tubules and stored for 4 weeks at -20°C. Avoid repeating freeze/thaw cycles.

4. **Reagent V:** Dissolved with 1 mL of distilled water before use. Mix thoroughly. It can be divided into small tubules and stored for 4 weeks at -20°C. Avoid repeating freeze/thaw cycles.

5. **Reagent VI:** Dissolved with 1 mL of distilled water before use. Mix thoroughly. It can be divided into small tubules and stored for 4 weeks at -20°C. Avoid repeating freeze/thaw cycles.

6. Working solution: Calculate according to the amount required for the experiment before use, according to the ratio of reagent I: reagent II: reagent III: reagent IV: distilled water=  $148\mu$ L:  $4\mu$ L:  $4\mu$ L:  $4\mu$ L:  $10\mu$ L(amount of 1T), mix well before use.

## **Product Description:**

Glycogen phosphorylase is divided into active glycogen phosphorylase a (Glycogen phosphorylase a, GPa) and inactive glycogen phosphorylase b (Glycogen phosphorylase b, GPb) two forms. The decomposition of glycogen is mainly carried out under the catalysis of glycogen phosphorylase a. When no activator is added, glycogen phosphorylase a catalyzes the production of glucose residues from glycogen and inorganic phosphorus to glycogen and glucose 1-phosphate. Under the action of phosphoglucose mutase and 6-phosphate glucose dehydrogenase, it further catalyzes the reduction of NADP to NADPH. Measuring the rate of increase of NADPH at 340nm can reflect the activity of glycogen phosphorylase a.

## **Reagents and Equipment Required but Not Provided:**

Ultraviolet spectrophotometer/microplate reader, low temperature centrifuge, constant temperature incubator/water bath, adjustable pipette, mortar/homogenizer, micro quartz cuvette/96 well UV plate, ice and distilled water.

# Procedure

# I. Sample preparation:

1. Tissue sample: according to the proportion of tissue weight (g): extraction solution volume (mL) of 1:5-10 to extract. It is suggested that 0.1 g of tissue with 1 mL of Extraction reagent and fully homogenized on ice bath. Centrifuge at 8000  $\times$ g for 10 minutes at 4°C to remove insoluble materials, and take the supernatant on ice before testing.

2. Bacteria or cells: collecting bacteria or cells into the centrifuge tube, suggested 5 million with 1 mL of Extraction reagent. Use ultrasonication to splitting bacteria and cells (placed on ice, ultrasonic power 200w, working time 3 seconds, interval 10 seconds, repeat for 30 times). Centrifuge at 8000  $\times$ g for 10 minutes at 4°C to remove insoluble materials, and take the supernatant on ice for testing.

3. Serum (plasma) sample: detect sample directly.

## II. Determination procedure:

1. Preheat ultraviolet spectrophotometer/microplate reader for 30 minutes, adjust wavelength to 340 nm, set zero with distilled water.

2. Working solution are preheated at 37°C for 5min.

3. GPa activity: add 10  $\mu$ L sample, 10  $\mu$ L reagent V, 10  $\mu$ L reagent VI, 170  $\mu$ L working solution in the micro quartz cuvette/96 well UV plate, mix immediately, and record the absorbance value at 10s at 340 nm A1, quickly place it in 37°C water bath or incubator (The microplate reader has a temperature control function that can adjust the temperature to 37°C) for 10 minutes, take it out and quickly dry it and measure the absorbance value A2 in 10min10s.Calculate  $\Delta A$ =A2-A1.Blank tube only need to be test one or two times.

## **III. Calculations:**

(1) Micro glass cuvette

A. Protein concentration

Unit definition: One unit of enzyme activity is defined as the amount of enzyme produce 1 nmol NADPH per minute every milligram protein in 37°C.

 $GPa \ (nmol/min/mg \ prot) = [\Delta A \times V_R \div \ (\epsilon \times d) \ \times 10^9] \div (V_S \times Cpr) \div T = 321.54 \times \Delta A \div Cpr$ 

B. Sample weight

Unit definition: One unit of enzyme activity is defined as the amount of enzyme produce 1 nmol NADPH per minute every gram tissue in 37°C.

 $GPa \ (nmol/min/g \ weight) = [\Delta A \times V_R \div \ (\epsilon \times d) \ \times 10^9] \div (W \times V_S \div V_E) \div T = 321.54 \times \Delta A \div W$ 

C. Sample volume

Unit definition: One unit of enzyme activity is defined as the amount of enzyme produce 1 nmol NADPH per minute every milliliter volume in 37°C.

GPa (nmol/min/mL) =[ $\Delta A \times V_R \div$  ( $\epsilon \times d$ ) ×10<sup>9</sup>] $\div V_S \div T=321.54 \times \Delta A$ 

D. Bacteria or cells number

Unit definition: One unit of enzyme activity is defined as the amount of enzyme produce 1 nmol NADPH per minute every million bacteria or cells in 37°C.

 $GPa \ (nmol/min/10^{4} cells) = [\Delta A \times V_{R} \div \ (\epsilon \times d) \ \times 10^{9}] \div (cell number \times V_{S} \div V_{E}) \div T = 321.54 \times \Delta A \div cell number \times V_{S} \div V_{E})$ 

 $\varepsilon$ : NADPH molar extinction coefficient, 6220 L/mol/cm;

d: Cuvette light path, 1cm;

V<sub>R</sub>: Total reaction volume, 0.2 mL;

V<sub>S</sub>: Add sample volume,0.01mL;

V<sub>E</sub>: extract volume, 1 mL;

W: Sample weight, g;

Cpr: Protein concentration of sample, mg/mL;

T: Reaction time, 10min

(2) 96-Well flat-bottom plates

Modify the d-1cm in the above formula to d-0.6cm (the light path of the 96-well plate) for calculation.

## Note:

1. If the measured absorbance value  $\Delta A$ >0.6, it is recommended to dilute the sample before measuring, and multiply the dilution factor in the calculation formula; if the measured absorbance value is low or close to the blank OD value, it is recommended to increase the sample volume before performing the measurement.

## **Experimental example**

1. Take 0.1 g of rabbit liver tissue, add 1 mL of extract, homogenize in an ice bath, centrifuge at 8000 ×g for 10 minutes at 4°C; take the supernatant and place on ice for testing. Use micro quartz cuvette to operate according to the determination steps, $\Delta A = A2 - A1 = 0.3472 - 0.2253 = 0.1219$ , according to the formula Calculated activity: